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Studies toward the discovery of the next generation of antidepressants. Part 5: 3,4-Dihydro-2H-benzo[1,4]oxazine derivatives with dual 5-HT $_{1A}$ receptor and serotonin transporter affinity

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Abstract—The design, synthesis, and structure–activity relationship of two novel classes of benzoxazine derivatives with dual selective serotonin reuptake inhibitors and 5-HT $_{1A}$ receptor activities are described. © 2005 Elsevier Ltd. All rights reserved.

Selective serotonin reuptake inhibitors (SSRIs) have achieved great success in treating depression and related illnesses. SSRIs work by blocking the neuronal reuptake of serotonin (5-HT), increasing the concentration of synaptic 5-HT and thus, increasing the activation of postsynaptic 5-HT receptors. Unfortunately, long-term treatment is required before clinical efficacy is achieved. This is speculated to be the result of acute stimulation of the somatodendritic 5-HT $_{1A}$ autoreceptors that decrease neuron firing to release serotonin in the forebrain. Only after desensitization of these autoreceptors, the serotonergic neurons resume normal firing and the eventual increase in 5-HT levels in the major forebrain areas. $^{1-3}$

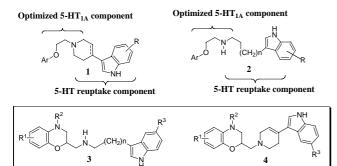
It has been proposed that addition of a 5-HT $_{1A}$ antagonist component to the action of an SSRI can limit the negative feedback through blockage of the autoreceptor, allowing an immediate increase in synaptic 5-HT. Preclinical evidence has shown that acute SSRI-induced increases in forebrain 5-HT can be significantly augmented by co-treatment with either pindolol or the more selective 5-HT $_{1A}$ antagonist WAY-100635.

more, co-administration of fluoxetine and WAY-100635 produces an immediate increase in 5-HT levels in rat frontal cortex using in vivo microdialysis, whereas this effect cannot be seen with fluoxetine treatment alone.⁶ In support of this hypothesis, clinical trials performed by Artigas et al.⁷ and Blier and Bergeron⁸ demonstrated that combination of (±)-pindolol, a nonselective 5-HT_{1A} antagonist, with the SSRI paroxetine shortened the onset of antidepressant action to a period of 3-7 days in contrast to the 2-3 weeks required with the SSRI alone. Therefore, incorporating both 5-HT_{1A} antagonism and 5-HT reuptake inhibition within a single molecule should provide an immediate increase in 5-HT in frontal cortex, resulting in a rapid onset antidepressant. This dual-action feature would thus form the basis of the next generation of antidepressant therapy.9-13

Previous reports from our laboratories have revealed two novel series capable of modulating both SSRI and 5-HT_{1A} receptor activities by linking an aryloxyethylamine with a 3-indoletetrahydropyridine (1) or a 3-indolealkylamine (2) through a common basic nitrogen. ^{12,13} In both series, aryloxyethylamines have been successfully utilized to produce potent 5-HT_{1A} ligands, while the indoletetrahydropyridine or 3-indolealkylamine moieties have served as robust 5-HT uptake inhibitor

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moieties. However, shortcomings were observed in both series, that is, lack of selectivity over the α_1 receptor. In order to expand upon the type of structures that can potentially maintain both desired activities, we decided to constrain the 5-HT_{1A} pharmacophore within a cyclic ring, benzoxazine ring, in a manner similar to that previously exemplified with benzodioxan moiety. 14 We are particularly interested in the benzoxazine system, since from a synthetic point of view, the benzoxazine analog offered the advantage of constraining the aryloxyethylamine moiety, as well as allowing us to probe another region in space via functionalization of the nitrogen of the oxazine ring. We now report the syntheses and the structure-activity relationships (SARs) of two new classes of benzoxazine 3-indolealkylamines 3a-q and benzoxazine 3-indoletetrahydropiridine analogs 4a-e.



Schemes 1–4 show the syntheses of target molecules 3a-q and 4a-e. The condensation of commercially available 2-amino-phenol and 2,3-dibromopropionic acid ethyl ester afforded ethyl 3,4-dihydro-2H-benzo[1,4]-oxazine-2-carboxylate 31 in moderate yield (33%). The nitrogen of the oxazine ring was alkylated under standard conditions and the ester reduced to generate benzoxazine derivative 32. Alcohols 32 were converted to the tosylates 33 using p-toluenesulfonyl chloride in pyridine, and coupling of tosylates 33 with different 3-indolepro-

Scheme 1. Reagents and conditions: (a) BrCH₂CHBrCO₂Et/K₂CO₃/ acetone, reflux; (b) R²Br or (R²I)/Et₃N/CH₃CN, reflux; (c) 1,4-*cyclo*-hexanedione/*p*-toluenesulfonic acid/toluene, reflux; (d) (R²CO)₂O/THF, reflux; (e) LiBH₄; (f) LiAlH₄/THF, rt; (g) TsCl/py, rt; (h) 3-indoealkylamines/Et₃N/DMSO, 90 °C.

Scheme 2. Reagent and conditions: (a) LiBH₄/THF, rt; (b) (*t*-Boc)₂O/THF, reflux; (c) BnBr/Et₃N/CH₃CN, rt; (d) TsCl/Py, rt; (e) NaN₃/DMF, 60 °C; (f) Ph₃P/THF-H₂O, rt; (g) LiAlH₄/THF, reflux; (h) 3-indolealkanoic acid/DMAPC/CH₂Cl₂, rt; (i) TFA/CH₂Cl₂, rt; (j) 5-fluoro-3-indolepropylbromide/Et₃N/DMSO, 90 °C.

MeO
$$\frac{1}{N}$$
 OEt $\frac{1}{N}$ MeO $\frac{1}{N}$ OH $\frac{1}{N}$ MeO $\frac{1}{N}$ OH $\frac{1}{N}$ MeO $\frac{1}{N}$ MeO

Scheme 3. Reagent and conditions: (a) LiBH₄/THF, rt; (b) (*t*-Boc)₂O/THF, reflux; (c) R²Br/Et₃N/CH₃CN, rt; (d) TsCl/Py, rt; (e) 5-fluoro-3-indolepropylamine/Et₃N/DMSO, 90 °C; (f) TFA/CH₂Cl₂, rt; (g) 5-fluoro-3-indolepyridine/Et₃N/DMSO, 90 °C.

Scheme 4. Reagent and conditions: (a) TsCl/Py, rt; (b) 3-indolepyridine/Et₃N/DMSO, 90 °C; (c) TFA/CH₂Cl₂, rt.

pylamines¹³ afforded the desired products $3\mathbf{d}$, \mathbf{f} - \mathbf{j} , \mathbf{n} , \mathbf{p} , and \mathbf{q} .

In Scheme 2, reduction of ester 31 with LiBH₄ led to alcohols 34, which were then treated either with ditert-butyldicarbonate or benzylbromide to give alcohols 35. Alcohols 35 were converted to amines 36 using a standard protocol. ¹³ Coupling of 36 to the appropriate 3-indolealkanoic acids, followed by reduction and deprotection, afforded target compounds 31,k, and m. Treatment of amines 36 with 5-fluoro-3-indolepropylbromide ¹³ resulted in target compounds 3c,e, and o.

In Scheme 3, reduction of 3-oxo-benzoxazine derivative 37 with LiBH₄ gave alcohol 38, which was then treated either with di-tert-butyldicarbonate or different halides to afford 39. The target compounds 3a and b were prepared by heating the corresponding tosylates 40 and 5-fluoro-3-indolepropylamine, followed by deprotection in TFA. Coupling the tosylates 40 with 5-fluoro-3-indolepyridine¹² gave target compounds **4a**–**c** and **e**.

The preparation of the benzoxazine analog 4d is shown in Scheme 4. Alkylation of tosylate 41 with 3-indole tetrahydropyridine¹² gave the desired product **4d** in good yield.

All compounds listed in Tables 1 and 2 were the racemates. 19 Serotonin transporter (r-5-HT-T) and 5-HT_{1A} receptor (h-5-HT_{1A}) and α_1 affinities, as well as 5- HT_{1A} intrinsic activity (GTP γ S E_{max}) are tabulated in Tables 1 and 2.

Table 1. Dual SSRI and 5-HT_{1A} receptor activities for compounds 3a-q^a

Compound	\mathbb{R}^1	\mathbb{R}^2	\mathbb{R}^3	n	r-5-HT-T K _i (nM) ^b	h-5-HT _{1A} K_i (nM) ^c	GTPS E _{max} (EC ₅₀ nM) ^d	$\alpha_1 K_i (nM)^e$
3a	8-OCH ₃	Н	F	1	3.13 ± 1.73	9.31 ± 1.25	66 ± 9.2 (52.2)	7.6 ± 3.6
3b	7 -OCH $_3$	H	F	1	11.70 ± 4.67	103.00 ± 15.34	$56 \pm 17.7 \ (181.5)$	44.5 ± 16.3
3c	Н	Me	F	1	12.43 ± 9.51	8.46 ± 2.27	$80 \pm 0.7 \ (167.5)$	26% at 100 nM
3d	Н	Et	F	1	15.28 ± 0.39	15.31 ± 2.20	$100 \pm 0 \ (90.0)$	_
3e	Н	H	F	1	15.98 ± 0.04	10.98 ± 3.14	$100 \pm 0 \ (155.0)$	70.0 ± 1.4
3f	Н	i-Pr	F	1	21.13 ± 5.48	2.81 ± 0.10	$90 \pm 7.1 \ (190.5)$	_
3g	Н	n-Pr	Н	1	22.73 ± 4.63	12.76 ± 2.84	$100 \pm 0 \ (196.5)$	481 ± 229.8
3h	H	i-Pr	H	2	23.78 ± 5.34	90.42 ± 29.12	$88 \pm 12.7 (329.0)$	_
3i	7-Cl	Me	F	1	42.58 ± 2.02	75.18 ± 20.45	$64 \pm 13.4 (336)$	607 ± 272.9
3j	Н	<i>i</i> -Bu	F	1	51.48 ± 47.41	8.15 ± 0.70	$88 \pm 12.7 (329)$	_
3k	Н	Me	H	1	88.50 ± 33.23	5.86 ± 1.12	$95 \pm 7.1 (33.6)$	28% at 100 nM
31	Н	H	Н	1	89.75 ± 15.20	6.91±0.52	$96 \pm 6.4 (93.5)$	26% at 100 nM
3m	Н	Bn	H	2	108.75 ± 39.24	76.14 ± 20.62	$64 \pm 7.8 \ (1385)$	_
3n	Н	Ph	F	1	43% at 100 nM	7.14 ± 0.22	_	65% at 100 nM
30	H	Bn	F	1	23% at 100 nM	7.25 ± 1.52	_	_
3p	Н	Ph	H	1	23% at 100 nM	30.21 ± 10.96	_	_
3q	H	Ph	Н	2	7% at 100 nM	75.77 ± 8.90	_	_

^a K_i, E_{max}, and EC₅₀ values are means of at least two experiments ± SEM (performed in triplicate, determined from nine concentrations).

Table 2. Dual SSRI and 5-HT_{1A} receptor activities for compounds $4a-e^a$

$$\mathbb{R}^1$$
 \mathbb{R}^2
 \mathbb{N}
 \mathbb{N}
 \mathbb{R}^3

Compound	\mathbb{R}^1	\mathbb{R}^2	\mathbb{R}^3	r-5-HT-T $K_i (nM)^b$	h-5-H T_{1A} K_i $(nM)^c$	GTPγS E_{max} (EC ₅₀ nM) ^d	$\alpha_1 K_i (nM)^e$
4a	8-OCH ₃	Н	F	6.94 ± 4.61	154.85 ± 3.15	$35 \pm 15.6 (386)$	14.5 ± 3.6
4b	8-OCH ₃	Me	F	12.58 ± 5.76	139.9 ± 8.56	$93 \pm 9.9 (190)$	96.5 ± 29.0
4c	8-OCH ₃	Et	F	13.15 ± 1.63	136.45 ± 18.00	$54 \pm 26.9 (418)$	229.0 ± 33.9
4d	Н	Н	Н	8.37 ± 10.79	804.80 ± 123.33	$62 \pm 0.7 (1131)$	16.6 ± 18.1
4e	8-OCH ₃	n-Pr	F	22.58 ± 3.64	362.80 ± 46.95	$92 \pm 0.7 (135)$	505.5 ± 222.7

^a K_i, E_{max}, and EC₅₀ values are means of at least two experiments ± SEM (performed in triplicate, determined from nine concentrations).

^b Binding affinity at rat cortical 5-HT reuptake sites labeled with [³H]-paroxetine. ¹⁵

^c Binding affinity at human 5-HT_{1A} receptors in CHO cells labeled with [³H]-8-OH-DPAT. ¹⁶

d Stimulation of [35 S]-GTP γ S binding in CHO cells expressing 5-HT_{1A} receptor; 17 E_{max} refers to maximal agonist effect relative to 5-HT. e Binding affinity at rat cortical al adrenergic receptors labeled with [3H]-prazosin. 18

^b Binding affinity at rat cortical 5-HT reuptake sites labeled with [³H]-paroxetine. ¹⁵

^c Binding affinity at human 5-HT_{1A} receptors in CHO cells labeled with [³H]-8-OH-DPAT. ¹⁶

^d Stimulation of [35 S]-GTPγS binding in CHO cells expressing the 5-HT_{1A} receptor; 17 E_{max} refers to maximal agonist effect relative to 5-HT. ^e Binding affinity at rat cortical al adrenergic receptors labeled with [3H]-prazosin. ¹⁸

Shown in Table 1 are the biological results of target molecules 3a-q. Consistent with our previous paper (Part 3),¹³ substitution of the 5-fluoro moieties to the indole ring resulted in a significant improvement in 5-HT transporter affinity but little effect on the affinity for 5-HT_{1A} receptor (3c vs k). Interestingly, incorporation of substitutents at the nitrogen of the oxazine ring had a minor effect on affinity for the 5-HT_{1A} receptor, indicating a large tolerance of the 5-HT_{1A} receptor toward structural variation in this region of the molecule. However, 5-HT transporter affinity dramatically decreased as the substitutent on the nitrogen of the oxazine ring increased in size (3c vs d,f,j,n, and o). A 5-fold increase in 5-HT transporter affinity was manifested when the methoxy group was attached at the 8-position (3a vs e). Surprisingly, when the methoxy group was moved to the 7-position, a 9-fold loss in affinity for 5-HT_{1A} receptor was found as well as ~4-fold loss in affinity for 5-HT transporter (3a vs b). In contrast to our previous studies (Parts 2 and 3), three compounds (3c,k, and 1) in this series had very low α_1 affinity. Unfortunately, all compounds from this study were found to be either 5-HT_{1A} agonists or partial agonists as measured by the GTP_{\gamma}S assay.

Table 2 depicts the investigation of the indole tetrahydropyridinyl group utilized as a transporter moiety in the benzoxazine series. Comparing compound 3a to 4a, a 16-fold loss in 5-HT_{1A} affinity was observed. A slight trend toward lower 5-HT transporter affinity was observed when the nitrogen of the oxazine ring was alkylated (4a-c and e). A 2-fold loss in affinity 5-HT_{1A} receptor was found when R² was an isopropyl group (4a vs e). Surprisingly, though we previously showed that compounds employing the tetrahydropyridinyl moiety had 5-HT_{1A} antagonism, 12 all the compounds listed in Table 2 were found to be either full or partial 5-HT_{1A} agonists in the GTP γ S assay in this report. In contrast to most of the compounds in Table 1, the benzoxazine indole tetrahydropyridinyl derivatives in Table 2 had higher affinity for α_1 receptor (i.e., **4a,b**, and **d** were more potent for α_1 receptor than the 5-H T_{1A} receptor).

In conclusion, we have discovered that the benzoxazine moiety can be utilized to embrace both the 5-HT $_{1A}$ pharmacophore along with the 5-HT transporter moiety to access dual SSRI and 5-HT $_{1A}$ receptor activities. The selectivity over α_1 receptor 20 was improved in several compounds, though most of the compounds in these two classes were found to function as 5-HT $_{1A}$ receptor agonists. Studies in our laboratories are continuing to further understand the 5-HT transporter and 5-HT $_{1A}$

pharmacophoric criteria within a single molecule and to identify novel agents that can represent the next generation of rapid onset antidepressants.

References and notes

- 1. Blier, P.; Bergerpm, R. J. Clin. Psychiat. 1998, 16.
- Kreiss, D. S.; Lucki, I. J. Pharmacol. Exp. Ther. 1995, 274, 219.
- 3. Hjorth, S.; Auerbach, S. B. Behav. Brain Res. 1996, 73, 281.
- 4. Romero, L.; Bel, N.; Casanovas, J. M.; Artigas, F. Int. Clin. Psychopharmacol. 1996, 11, 1.
- Hjorth, S.; Westlin, D.; Bengtsson, H. J. Neuropharmacology 1997, 36, 461.
- 6. Dawson, L. A.; Nguyen, D. L.; Schechter, L. E. *J. Psychopharacol.* **2002**, *16*, 145.
- 7. Artigas, F.; Perez, V.; Alvarez, E. Arch. Gen. Psychiat. 1994, 51, 248.
- 8. Blier, P.; Bergeron, R. J. Clin. Psychopharmacol. 1995, 15, 217
- Martinez-Esparza, J.; Oficialdegui, A. M.; Perez-Silanes, S.; Heras, B.; Orus, L.; Palop, J. A.; Lasheras, B.; Roca, J.; Mourelle, M.; Bosch, A.; Del Castillo, J. C.; Tordera, R.; Del Rio, J.; Monge, A. J. Med. Chem. 2001, 44, 418.
- Takeuchi, K.; kohn, T. J.; Honigschmidt, N. A.; Rocco, V. P.; Spinazze, P. G.; Koch, D. J.; Atkinsion, S. T.; Hertel, L. W.; Nelson, D. L.; Wainscott, D. B.; Ahmad, L. J.; Shaw, J.; Threlkeld, P. G.; Wong, D. T. *Bioorg. Med. Chem. Lett.* 2003, 13, 2393.
- Rocco, V. P.; Spinazze, P. G.; Kohn, T. J.; Honigschmidt, N. A.; Nelson, D. L.; Wainscott, D. B.; Ahmad, L. J.; Shaw, J.; Threlkeld, P. G.; Wong, D. T.; Takeuchi, K. Bioorg. Med. Chem. 2004, 14, 2653.
- Mewshaw, R. E.; Meagher, K. L.; Zhou, P.; Zhou, D.; Shi, X.; Scerni, R.; Smith, D. L.; Schechter, L. E.; Andree, T. H. Bioorg. Med. Chem. Lett. 2002, 12, 307.
- Mewshaw, R. E.; Zhou, D.; Zhou, P.; Shi, X.; Hornby, G.;
 Spangler, T.; Scerni, R.; Smith, D.; Schechter, L. E.;
 Andree, T. H. J. Med. Chem. 2004, 47, 3823.
- Kang, Y.; Stack, G. P. U.S. Patent 5,750,724, 1998;
 Husbands, G. M.; Stack, G, P.; Mewshaw, R. E. U.S. Patent 6,559,169, 2003.
- Cheetham, S. C.; Viggers, J. A.; Slater, N. A.; Heal, D. J.; Buckett, W. R. Neuropharmacology 1993, 32, 737.
- Dunlop, J.; Zhang, Y.; Smith, D. L.; Schechter, L. E. J. Pharmacol. Toxicol. Methods 1998, 40, 47.
- Larenzo, S.; Birdsall, N. J. Br. J. Pharmacol. 1993, 109, 1120.
- 18. Morrow, A.; Creese, I. Mol. Pharmacol. 1986, 29, 321.
- For experimental procedures, see: US patents: Mewshaw,
 R. E.; Zhou, D. U.S. Patent 6,221,863, 2001; Mewshaw,
 R. E.; Shaw, U. U.S. Patent 6,313,114, 2001.
- Peroutka, S. J.; U'Prichard, D. C.; Greenberg, D. A.;
 Snyder, S. H. Neuropharmacology 1977, 16, 549.